DNA Logic Circuits with a DNA Polymerase and a Nicking Enzyme

Ryo Hirose¹, Satoshi Kobayashi¹, Ken Komiya²

Department of Communication Engineering and Informatics
 Graduate School of Informatics and Engineering
 University of Electro-Communications, Tokyo, Japan
Department of Computational Intelligence and Systems Science
 Interdisciplinary Graduate School of Science and Engineering
 Tokyo Institute of Technology, Yokohama, Japan

Abstract—The current most popular and established approach to DNA logic circuits is the implementation by DNA strands reaction networks where toehold-mediated strand displacement invokes and performs the evaluation of each DNA logic gate. Strand displacement approach requires, however, a large amount of time, for instance, approximately 30 minutes, to execute a logic operation ([18]). Furthermore, the concentration of an output molecule released from a gate can not exceed that of the gate molecule. Therefore, it is often the case that large quantities of input and gate molecules are required when the gate is of large out-degree. In order to overcome these problems, it is indispensable to devise a DNA logic gate which runs quickly and can amplify the quantity of the output molecule. We will propose a DNA implementation of logic gates with such good properties using DNA polymerase and nicking enzyme.

Keywords: DNA Computing, Logic Circuit, DNA Polymerase, Nicking Enzyme

1. Introduction

In 1994, Adleman initiated the paradigm of DNA computing by devising and demonstrating a biological experimental protocol to solve Directed Hamiltonian Path Problem([1]). It has achieved important progress of the technologies in making DNAs work as computing devices([4][3][2], etc.). They also produce some new and important key technologies([14][10], etc.) in the field of DNA nanotechnology, where it is aimed to construct intended nano-scale shapes or structures by self-assembly of DNA molecules. In these fields, it is currently emerging the movement for establishing the methodology to construct automatic molecular robots performing intended tasks in some specified environment ([8][5],etc.). A molecular robot should, however, contain at least three important components, a sensor, a circuit, and an actuator, in itself. The current technology ([8][5]) does not satisfy these three requirements, and it is still at a premature stage. So, it is very challenging to explore a possible framework of the methodology to construct a molecular robot with a sensor, a circuit, and an actuator.

Molecular circuits equipped in molecular robots should serve as logic circuits, memory devices, and control devices, etc ([12]). In this paper, we will focus on a molecular circuit as a computing device performing logic operations. There are many works which have proposed DNA implementations of logic circuits. One of the most popular and established approach to DNA logic circuits is the implementation by DNA strands reaction networks where toehold-mediated strand displacement ([16]), strand displacement for short, invokes and performs the evaluation of each DNA logic gate([17][15][18]). Strand displacement approach requires, however, a large amount of time, for instance, approximately 30 minutes, to execute a logic operation([18]). Furthermore, the concentration of an output molecule released from a gate can not exceed that of the gate molecule. Therefore, it is often the case that large quantities of input and gate molecules are required when the gate is of large out-degree. In order to overcome these problems, it is indispensable to devise a DNA logic gate which runs quickly and can amplify the quantity of the output molecule. We will propose a DNA implementation of logic gates with such good properties using a DNA polymerase and a nicking enzyme.

2. Preliminaries

DNA is a molecule consisting of simple units, called *nucleotides*, with backbones made of sugars and phosphate groups joined by phosphodiester bonds. Each nucleotide contains one of four types of atomic groups, called bases, each being either of adenine (abbreviated A), cytosine (C), guanine (G) and thymine (T). These long polymers are more commonly called *strands*, and short polymers are called *oligonucleotides*, or simply *oligos*. Note that every DNA strand has two distinct ends, one with a free 5' phosphate group and the other with a free 3' hydroxyl group, referred to as the 5' and 3' ends, respectively. Thus, we can regard a sequence of nutleotides as having a natural orientation from its 5' to 3' ends, and it is often written as a sequence of letters (bases) A, C, G, T oriented from 5' to 3' direction.

Under some appropriate chemical conditions, two strands will pair up and twist around each other to form a fa-

mous double helix structure discovered by Watson and Crick. The pairing happens between A's and T's and between G's and C's, which is called Watson-Crick base pairing. This pairing occur only if the two strands runs in an antiparallel fashion. For instance, the strands 5'-GCATCAG-3' and 5'-CTGATGC-3'will form a hybridized object $\frac{5'-GCATCAG-3'}{3'-CGTAGTC-5'}$. These kinds of completely hybridized DNA strands are called *double strands*. A strand with no base pairing with other strands is called a *single strand*.

DNA polymerase is an enzyme, or molecular machine, which reads a single strand X in the 3' to 5' direction, and builds the complementary strand X^* in the 5' to 3' direction, one nucleotide at a time, where the strand X is called a template. The activation of DNA polymerase to work as a (complementary) copy machine requires a short portion of double stranded part in the template. That is, we need a short piece of single strand, called *primer*, which is complementary to some part of the template. It is onto the 3' end of this primer that DNA polymerase will add new nucleotides. So, in order to make the copy X^* of X, we need a primer which is complementary to the last 3' end portion of the strand X.

3. Related Work

The current most promising approach to the construction of DNA logic gates is the DNA implementation of logic operation by the use of strand displacement. Its basic mechanism is illustrated in Figure 1. Consider a complex consisting of strands 5' - A - B - 3' and $5' - C^* - B^* - 3'$ with B and B* hybridized, where A, B, C are sequences of bases. If another strand 5' - B - C - 3' exists in a solution, then 5' - B - C - 3' hybridizes to the complex with C and C* hybridized. The strand 5' - B - C - 3' gradually extends its pairing with $5' - C^* - B^* - 3'$ by replacing the strand

5' - A - B - 3' in a random walk fashion. This process is called "branch migration".

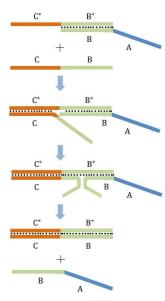


Fig. 1: Toehold-mediated Strand Displacement

There have been many research works to propose biolab methods to implement some computational tasks by the use of strand displacement ([11][17][15][18],etc.), and it is considered as one of the most promising bio-lab techniques in DNA computing. The computational capability of strand displacement is also an interesting research topic from the viewpoint of computation theory([12][9],etc.).

Strand displacement is, however, a slow reaction if we want to use it as an essential bio-lab operation to implement DNA logic circuits of molecular robots. Furthermore, in these approaches, the concentration of an output molecule released from a gate can not exceed that of the gate molecule. In this sense, the construction of large circuits based only on strand displacement is not feasible.

In the next section, we will propose a new bio-lab method for constructing logic circuits with DNA molecules, where we use the amplification system based on nicking enzyme and DNA polymerase. Waker first applied this reaction to the construction of isothermal amplification system of DNA ([13]). The idea of using nicking enzyme to computational molecular devices was also accomplished by Matsuda and Yamamura ([6]), where they used a DNA polymerase and a nicking enzyme in order to cascade molecular state transition systems based on Whiplash PCR([3]). Very recently, Montagne, et al., proposed to use a DNA polymerase and a nicking enzyme to construct a programmed DNA oscillator([7]). In this paper, we will apply a similar reaction system to the construction of DNA logic circuits, and we will show by computational simulations that the proposed system is more efficient than the strand displacement approaches. Furthermore, in the proposed system, the output of each DNA logic gate can be amplified by the use of a DNA polymerase and a nicking enzyme.

4. Construction of Combinatorial Circuit

In this section, we will propose a method to construct combinatorial circuits by using DNA polymerase and nicking enzyme. As in other related works about DNA logic gates, we will prepare for each boolean variable x, a DNA strand X. The existence of X in the solution implies the evaluation of the boolean variable x as 1. On the other hand, how can we encode the evaluation x=0 into the solution? For this purpose, we will also prepare another DNA strand NX, and we regard the existence of NX in the solution as the evaluation of the variable x as 0. Thus, the strand X and X can not exist at the same time in the solution. Although most of the other works encode the evaluation X in the nonexistence of X in the solution, we will use the negation strand X in order to implement NOT gate simply.

In order to implement DNA logic circuits, we will use the following three basic *abstract-level* chemical reactions:

- AND reaction: A∧B → C the strand C is produced if and only if both of the strands A and B exist in the solution.
- 2. OR reaction A | B \rightarrow C the strand C is produced if and only if either of the strands A or B exists in the solution.
- 3. PROPAGATE (PROP) reaction : $A \rightarrow B$ the strand B is produced if and only if the strand A exists in the solution.

When we want to construct a chemical reaction system which utilizes any given boolean function, it suffices to devise chemical implementation of logic gates, AND, OR, and NOT. Then, it is easy to construct AND, OR, and NOT gates using the three basic reactions above.

- AND gate construction Consider an AND gate with input variables a and b and with an output variable c. Then, we will prepare the strands A, NA for the variable a, B, NB for the variable b, C, NC for the variable c. It is easy to see that the AND and OR reactions, A ∧ B → C and NA | NB → NC, implement the AND gate.
- 2. OR gate construction Consider an OR gate with input variables a and b and with an output variable c. Then, we will prepare the strands A, NA for the variable a, B, NB for the variable b, C, NC for the variable c. It is easy to see that the AND and OR reactions, $A \mid B \rightarrow C$ and $NA \land NB \rightarrow NC$, implement the OR gate.
- 3. NOT gate construction Consider a NOT gate with an input variable a and with an output variable b. Then, we will prepare the strands A, NA for the variable a,

 $B,\ NB$ for the variable b. It is easy to see that the PROP reactions, $A\to NB$ and $NA\to B$, implement the NOT gate.

In the rest of this section, we will propose a method to implement AND, OR and PROP reactions with the use of DNA polymerase and nicking enzyme. We assume that we will use a nicking enzyme which recognizes a double stranded DNA sequences $3' - R - 3' \over 3' - R^* - 5'$, and cleave the 3' end of the lower strand R* only.

4.1 AND reaction: $A \wedge B \rightarrow C$

We will explain the construction of AND reaction which, with the existence of sufficient amounts of input DNA single strands A and B in a solution, outputs a single stranded DNA sequence C. Principal molecule of this re-

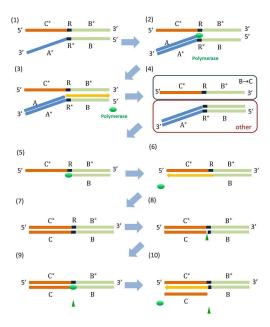


Fig. 2: AND Reaction

action, called *AND complex*, consists of two DNA strands $5' - C^* - R - B^* - 3'$ and $5' - B - R^* - A^* - 3'$ with B and B* hybridized at its initial state (Fig.2(1)). Let us consider the situation where sufficient amounts of DNA strands A and B exist in a solution. At first, A hybridizes to A* of the AND complex (Fig.2(2)). Then, DNA polymerase bind to this double stranded part (consisting of A and A*) and elongates the sequence A until it reaches to the 5'-end of the sequence $5' - B - R^* - A^* - 3'$ (Fig.2(3)). This makes the double stranded part (consisting of B and B*) detached from the AND complex and we will have DNA strand $5' - C^* - R - B^* - 3'$ in the solution (Fig.2(4)). Then, the strand B hybridizes to B* of $5' - C^* - R - B^* - 3'$ (Fig.2(5)), and DNA polymerase binds to this double stranded part (consisting of B and B*) and elongates the

sequence B until it reaches to the 5'-end of the sequence $5'-C^*-R-B^*-3'$ (Fig.2(6)). This complex is recognized by a nicking enzyme at the site R and R*, and it cleaves the strand R* (Fig.2(8)) . DNA polymerase, while pushing the strand C away, elongates the strand $5'-B-R^*-3'$ again until it reaches to the 5'-end of the sequence $5'-C^*-R-B^*-3'$ and the output strand C is released. Repeating the process (8), (9) and (10), the output strand C is amplified and released.

4.2 OR reaction — $A \mid B \rightarrow C$

We will explain the construction of OR reaction which, with the existence of sufficient amounts of input DNA single strands A or B in a solution, outputs a single stranded DNA sequence C. Principal molecule of this re-

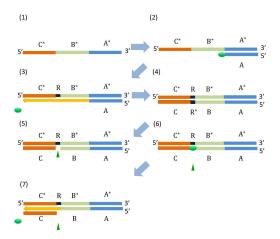


Fig. 3: OR Reaction

action, called OR complex, consists of single DNA strand $5' - C^* - R - B^* - A^* - 3'$ at its initial state (Fig.3(1)). Let us consider the situation where sufficient amounts of DNA strands A or B exist in a solution. At first, A or B hybridizes to A^* or B^* of the OR complex (Fig.3(2)). Fig.3 describes the case of A hybridizing to A*. Then, DNA polymerase binds to this double stranded part and elongates the sequence A until it reaches to the 5'-end of the sequence $5' - C^* - R - B^* - A^* - 3'$ (Fig.3(3),(4)). Next, a nicking enzyme recognizes R and R* and cleaves the strand R* (Fig.3(5)). DNA polymerase, while pushing the strand C away, elongates the strand $5'A - B - R^* - 3'$ again until it reaches to the 5'-end of the sequence $5' - C^* - R - B^* - A^* - 3'$ and the output strand C is released. Repeating the process (5), (6) and (7), the output strand C is amplified and released.

4.3 PROP reaction : $A \rightarrow B$

In this subsection, we will explain the construction of PROP reaction which, with the existence of sufficient amounts of input DNA single strands A in a solution, outputs a single stranded DNA sequence B.

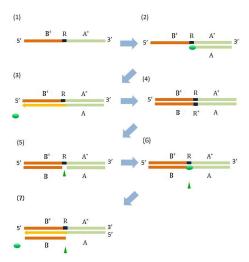


Fig. 4: PROP Reaction

Principal molecule of PROP reaction, called PROP complex, consists of single DNA strand $5' - B^* - R - A^* - 3'$ at its initial state (Fig.4(1)). Let us consider the situation where sufficient amounts of DNA strands A exist in a solution. At first, A hybridizes to A^* of the PROP complex (Fig.4(2)). Then, DNA polymerase binds to this double stranded part and elongates the sequence A until it reaches to the 5'-end of the sequence $5' - B^* - R - A^* - 3'$ (Fig.4(3),(4)). Next, a nicking enzyme recognizes R and R^* and cleaves the strand R^* (Fig.4(5)). DNA polymerase, while pushing the strand B away, elongates the strand $S' - A - R^* - S'$ again until it reaches to the 5'-end of the sequence $S' - B^* - R - A^* - S'$ and the output strand B is released. Repeating the process (5), (6) and (7), the output strand B is amplified and released.

5. Mathematical Model of Chemical Reaction Networks

In this section, we will explain the mathematical model of chemical reaction networks for the proposed combinatorial DNA circuits. We only give the explanation for the case of AND reaction, since the models of other basic operations, OR and PROP, are almost similar to that of AND reaction and can be obtained easily.

5.1 Chemical Reaction Network of AND Reaction

For any rate constant k, by k' we denote the rate constant of the reverse reaction of the reaction corresponding to k. Parameters k_{poly}^A , k_{poly}^B and k_{poly}^C are rate constants of polymerase reaction elongating the strands, A, B, and

C, respectively. The parameter k_{nick} is the rate constant for nicking enzyme to cleave strands.

$$\begin{aligned} \mathbf{G} + \mathbf{A} & \overset{k_A}{\rightleftharpoons} & \mathbf{G} \cdot \mathbf{A} \\ k'_A & & \\ \mathbf{G} \cdot \mathbf{A} & \overset{k_B^A}{\rightleftharpoons} & \mathbf{G}_1 + \mathbf{G}_2 \cdot \mathbf{A} \\ \mathbf{G}_1 + \mathbf{B} & \overset{k_B}{\rightleftharpoons} & \mathbf{G}_1 \cdot \mathbf{B} \\ \mathbf{G}_1 \cdot \mathbf{B} & \overset{k_{poly}}{\rightleftharpoons} & \mathbf{G}_1 \cdot \mathbf{B} \mathbf{C} \\ \mathbf{G}_1 \cdot \mathbf{B} & \overset{k_{nick}}{\rightleftharpoons} & \mathbf{G}_1 \cdot \mathbf{B} \cdot \mathbf{C} \\ \mathbf{G}_1 \cdot \mathbf{B} \cdot \mathbf{C} & \overset{k_{poly}^C}{\rightleftharpoons} & \mathbf{G}_1 \cdot \mathbf{B} \mathbf{C} + \mathbf{C} \end{aligned}$$

Fig. 5: Full Reaction Network of AND Gate

Chemical reaction network of an *abstract-level* AND reaction is described in Figure 5, where AND complex, its upper strand, and its lower strand are represented as G, G_1 , and G_2 , respectively.

5.2 Differential Equations of AND Reaction

Differential equation system to mathematically model and simulate the AND reaction is given in Figure 6.

6. Simulation Results

We used the following kinetic parameters for the simulation, which are determined by referring to the data reported in [7].

$$\begin{array}{ll} k_A = k_B = 2.4 \times 10^7 & (\mathrm{M}^{-1}\mathrm{min}^{-1}) \\ k_A' = k_B' = 1.2 \times 10^{-3} & (\mathrm{M}^{-1}\mathrm{min}^{-1}) \\ k_{Poly}^B = 2.4 \times 10^1 & (\mathrm{min}^{-1}) \\ k_{Poly}^A = k_{Poly}^C = 2.4 \times 10^1 \times 0.8 & (\mathrm{min}^{-1}) \\ k_{nick}^A = 6.0 \times 10^0 & (\mathrm{min}^{-1}) \end{array}$$

6.1 Simple Amplification Network

We first constructed a simple amplification network in Figure 7, whose experimental data were reported in [7], and simulated its behavior. The result is given in Figure 8, where we plot the concentration of the hybridized compound of the input strand A and the PROP complex molecule. The obtained simulation curve approximately coincides with the experimentally obtained fluorecense curve reported in [7], and the saturation time is approximately 4 - 6 minutes in both of the simulation and the experimental data. So, we can conclude that the mathematical model and its parameters given above work well corresponding to the experimental data.

$$\begin{split} \frac{d[A]}{dt} &= -k_A[G][A] + k_A'[G \cdot A] \\ \frac{d[B]}{dt} &= -k_B[G_1][B] + k_B'[G_1 \cdot B] \\ \frac{d[C]}{dt} &= k_{\mathrm{poly}}^{\mathrm{C}}[G_1 \cdot B \cdot C] \\ \frac{d[G]}{dt} &= -k_A[G][A] + k_A'[G \cdot A] \\ \frac{d[G \cdot A]}{dt} &= k_A[G][A] - k_A'[G \cdot A] - k_{\mathrm{poly}}^{\mathrm{A}}[G \cdot A] \\ \frac{d[G_1]}{dt} &= k_{\mathrm{poly}}^{\mathrm{A}}[G \cdot A] - k_B[G_1][B] + k_B'[G_1 \cdot B] \\ \frac{d[G_2 \cdot A]}{dt} &= k_{\mathrm{poly}}^{\mathrm{A}}[G \cdot A] \\ \frac{d[G_1 \cdot B]}{dt} &= k_B[G_1][B] - k_B'[G_1 \cdot B] - k_{\mathrm{poly}}^{\mathrm{B}}[G_1 \cdot B] \\ \frac{d[G_1 \cdot BC]}{dt} &= k_{\mathrm{poly}}^{\mathrm{B}}[G_1 \cdot B] - k_{\mathrm{nick}}[G_1 \cdot BC] + \\ &+ k_{\mathrm{poly}}^{\mathrm{C}}[G_1 \cdot B \cdot C] - k_{\mathrm{poly}}^{\mathrm{C}}[G_1 \cdot B \cdot C] \\ \frac{d[G_1 \cdot B \cdot C]}{dt} &= k_{\mathrm{nick}}[G_1 \cdot BC] - k_{\mathrm{poly}}^{\mathrm{C}}[G_1 \cdot B \cdot C] \end{split}$$

Fig. 6: Full system of differential equations for AND reaction

6.2 Majority Vote Circuit

We applied the proposed method to the construction of a majority vote circuit. The circuit outputs 1 if the number of input variables assigned to 1 is greater than that of input variables assigned to 0. We applied the method to 3-variable case. The circuit is given in Figure 9. For each variable, x, y, z, a, b, c, d, e, we will prepare the DNA strands X, Y, Z, A, B, C, D, E, for representing the evaluation of each variable to 1. Furthermore, the DNA strands NX, NY, NZ, NA, NB, NC, ND, NE are used for representing the evaluation of each variable to 0.

We set the concentration of each input strand X, Y, Z, NX, NY, NZ, to 1.0×10^{-10} (M) if it should exist in the solution, to 0 (M) otherwise. Furthermore, the concentration of the AND-complex and OR-complex at the 1st, 2nd, and 3rd layers are set to 1.0×10^{-9} (M), 1.0×10^{-7} (M), and 5.0×10^{-8} (M), respectively.

Figure 10 gives the simulation result for the case of inputs x=y=z=1. That is, we put 1.0×10^{-10} (M) of X,Y,Z, and 0 (M) of NX,NY,NZ, in the solution. As expected, the concentration of the output strand E grows rapidly, and NE stays at 0 for all the time of the simulation, which correctly simulates the behavior of the majority vote circuit.

For the case of inputs x=y=z=0. That is, we put 1.0×10^{-10} (M) of NX, NY, NZ, and 0 (M) of X, Y, Z, in the solution. As expected, the concentration of the output strand NE grows rapidly, and E stays at 0 for all the time

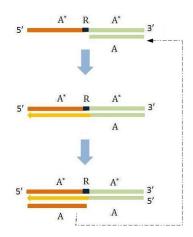


Fig. 7: Simple Amplification Network

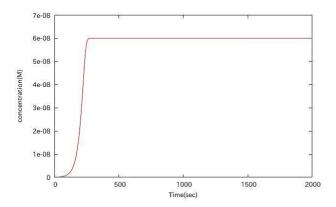


Fig. 8: Simulation of Simple Amplification Network

of the simulation, which correctly simulates the behavior of the majority vote circuit (Fig.11). Note that in Fig. 11, the concentrations of NA and NC are very small, so they almost stay at 0 in the graph.

Figure 12 gives the global view of the simulation result for the case of inputs x=y=z=1. We can verify that the growth of the 3rd layer output is very rapid (at most 10 minutes are enough for 3 steps of logic operation).

On the other hand, the output strands behavior largely depends on the initial concentrations of principal complex of each gate and each input strand. For instance, if we put 1.0×10^{-10} (M) of the input strands, 1.0×10^{-11} (M) of AND-complex and OR-complex in the solution, then, for the inputs x=y=z=1, the output strand E grows very slowly (See Figure 13). So, design of these concentration parameters are important problem in the future.

7. Conclusions

We proposed a new bio-lab method for constructing logic circuits with DNA molecules, where we use the amplification

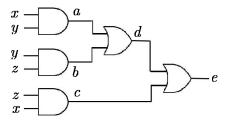


Fig. 9: Majority Vote Circuit

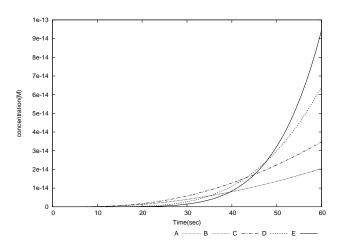


Fig. 10: Simulation of Majority Vote Circuit (1)

system based on a nicking enzyme and a DNA polymerase similar to [13], [6], and [7]. The proposed method has some good properties as computational devices that it is more time-efficient than the strand displacement approaches ([17][15][18], etc.) and that the output of each DNA logic gate can be amplified, and so, the scalability and the feasibility of the proposed system is better than the previous methods.

However, in order to increase the amount of the output molecule, the concentration of each gate complex should be carefully designed as is discussed in section 6.2. Furthermore, the proposed method has a problem that the amplification of the output strands continues until DNA polymerase uses up substrates. So, we need a bio-lab method for stopping or inhibiting the amplification process in this framework. An idea is to use inhibitor strands and exonuclease as in [7]. All of these issues are our future research topics.

Acknowledgement

The second author is supported in part by Grant-in-Aid for Scientific Research (C) No.22500010, Japan Society for the Promotion of Science.

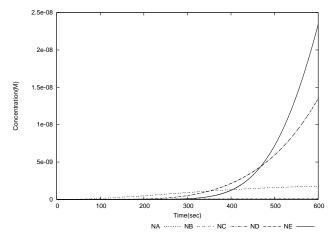


Fig. 11: Simulation of Majority Vote Circuit (2)

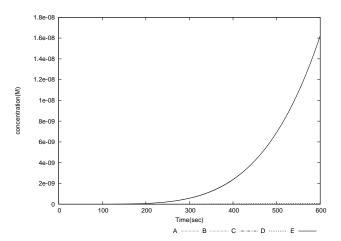


Fig. 12: Simulation of Majority Vote Circuit (3)

References

- [1] L. Adleman, Molecular Computation of Solutions to Combinatorial Problems. *Science* **266**, pp.1021-1024, 1994.
- [2] Y. Benenson, T. Paz-Elizur, R. Adar, E. Keinan, Z. Livneh, E. Shapiro, Programmable and autonomous computingmachine made of biomolecules, *Nature* 414, pp.430-434, 2001.
- [3] M. Hagiya, M. Arita, D. Kiga, K. Sakamoto, S. Yokoyama, Towards parallel evaluation of Boolean μ-formulas with molecules, In DNA Based Computers III(American Mathematical Society), pp.57-72, 1999.
- [4] R. Lipton, DNA solution of hard computational problem, *Science* 268, pp.542-545, 1995.
- [5] D. Zhang, E. Winfree, K. Lund, A. Manzo, N. Dabby, N. Michelotti, A. Johnson-Buck, J. Nangreave, S. Taylor, R. Pei, M. Stojanovic, N. Walter, E. Winfree, H. Yan, Molecular robots guided by prescriptive landscapes, *Nature*, 465, pp.206-210, 2010.
- [6] D. Matsuda, M. Yamamura, Cascading Whiplash PCR with a Nicking Enzyme, DNA Computing — Proc. of International Workshop on DNA-Based Computers, DNA8 (2002), Lecture Notes in Computer Science, pp.38-46, 2003.
- [7] K. Montagne, R. Plasson, Y. Sakai, T. Fujii, Y. Rondelez, Programming an in vitro RNA oscillator using a molecular networking strategy, Molecular Systems Biology, 7, Article Number 466, 2011.
- [8] T. Omabegho, R. Sha, N. Seeman, A Bipedal DNA Brownian Motor with Coordinated Legs, *Science* 324, pp.67-71, 2009.

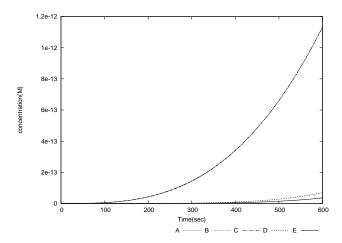


Fig. 13: Simulation of Majority Vote Circuit (4)

- [9] L. Qian, D. Soloveichik, E. Winfree, Efficient Turing-universal computation with DNA polymers, *Lecture Notes in Computer Science* 6518, pp.123-140, 2011.
- [10] P. Rothemund, Folding DNA to create nanoscale shapes and patterns, *Nature* 440, pp.297-302, 2006.
- [11] G. Seelig, D. Soloveichik, D. Zhang, E. Winfree, Enzyme-free nucleic acid logic circuits, *Science*, **314**, pp.1585-1588, 2006.
- [12] D. Soloveichik, G. Seelig, E. Winfree, DNA as a Universal Substrate for Chemical Kinetics, *Proc. Natl. Acad. Sci. USA*, **107**, pp.5393-5398, 2010
- [13] G. Walker, M. Little, J. Nadeau, D. Shank, Isothermal in vitro amplification of DNA by a restriction enzyme/DNA polymerase system, *Proc. Natl. Acad. Sci. USA*, 89, pp.392-396, 1992.
- [14] E. Winfree, F. Liu, L. Wenzler, N. Seeman, Design and self-assembly of two-dimensional DNA crystals, *Nature* 394, pp.539-544, 1998.
- [15] P. Yin, H. Choi, C. Calvert, N. Pierce, Programming biomolecular self-assembly pathways, *Nature*, 451, pp.318-322, 2008.
- [16] B. Yurke, A.J. Turberfield, A.P. Mills, F.C. Simmel, J. Neumann, A DNA-fuelled molecular machine made of DNA. Nature 406, pp.605-608, 2000.
- [17] D. Zhang, A. Turberfield, B. Yurke, E. Winfree, Engineering Entropy-Driven Reactions and Networks Catalyzed by DNA, *Science*, 318, pp.1121-1125, 2007.
- [18] D. Zhang, E. Winfree, Control of DNA Strand Displacement Kinetics Using Toehold Exchange, J. of American Chemical Society, 131, pp.17303-17314, 2009.